Diadenosine Polyphosphates in Tears of Sjögren Syndrome Patients

Gonzalo Carracedo, Assumpta Peral, and Jesús Pintor

Purpose. To analyze the levels of diadenosine tetraphosphate (Ap₄A) and diadenosine pentaphosphate (Ap₅A) in tears of subjects with Sjögren syndrome and to compare them with those in a control group.

Methods. Twelve subjects with a diagnosis of Sjögren syndrome and 20 healthy control subjects were invited to participate in the present study. Schirmer strips were used to measure tear secretion (Schirmer I test) and to collect tears. Ap₄A and Ap₅A were measured by high-pressure liquid chromatography (HPLC), and a dry eye questionnaire (DEQ) was used to evaluate dry eye symptomatology.

Results. The mean concentrations of Ap₄A and Ap₅A in the Sjögren syndrome group were 2.54 ± 1.02 and 26.13 ± 6.95 μM, respectively. This group of patients was divided in two subgroups: four patients with normal tear production and eight patients with low tear production. Concentrations of Ap₄A, and Ap₅A in patients with normal tear production (Schirmer test result, 12.3 ± 1.2 mm) were 0.47 ± 0.20 and 8.05 ± 3.27 μM, respectively. In the patients with low tear production (Schirmer test result, 1.0 ± 0.5 mm), the concentrations were 4.09 ± 1.36 and 39.51 ± 8.46 μM, respectively and in the control group, 0.13 ± 0.03 and 0.04 ± 0.02 μM, respectively.

Conclusions. Patients with Sjögren syndrome have abnormally elevated concentrations of diadenosine polyphosphates, indicating that these compounds could be used in the diagnosis of this disease. (Invest Ophthal Vis Sci. 2010;51:5452–5459)

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Sjögren’s syndrome is a chronic autoimmune disease in which the body’s immune system mistakenly attacks its own exocrine glands, particularly the salivary and the main lacrimal glands. Lymphocytes infiltrate the glands and cause a decrease in their secretions. This syndrome is classified as primary when only the exocrine glands are affected and as secondary when it is associated with other systemic diseases. Patients with Sjögren syndrome have abnormally increased concentrations of diadenosine polyphosphates in the tear film, which was described for the first time in 2002. These compounds Ap₄A, Ap₅A, and Ap₆A in rabbits, can stimulate tear secretion, giving these compounds a secretagogic action. Concerning the ocular surface, Ap₅A improves the rate of wound healing in the cornea of New Zealand White rabbits. Also, intraocular pressure (IOP) can be modulated by the action of some dinucleotides in rabbits.

Primary Sjögren syndrome has a prevalence rate of 0.5% and occurs more frequently in women than in men (9:1). It is diagnosed late, usually at 45 to 50 years of age, most often in an ophthalmologist’s or optometrist’s consulting room.

Methods

The study was conducted in compliance with good clinical practice guidelines, institutional review board regulations, informed consent regulations, and the tenets of the Declaration of Helsinki (World Medical Association, 2008). All the subjects enrolled in the study were adults older than 18 years and were able to give informed consent.

Subjects

Twelve subjects, 9 women and 3 men aged between 35 and 66 years (mean, 52.8 ± 4.2), who had received a diagnosis of primary Sjögren syndrome in the past 5 years (mean, 3.4 ± 0.9) and were enrolled in...
the Spanish Sjögren syndrome Society, participated voluntarily in the study. In all the patients, the disease was diagnosed according to the American-European Consensus Group (AECG) on diagnostic criteria for Sjögren syndrome (Vitali et al.2). This Sjögren syndrome group was divided into two subgroups, patients who had a Schirmer test result of more than 5 mm (Sjögren syndrome normal tear subgroup) and patients who had results below 5 mm (Sjögren syndrome low tear subgroup).

The control group constituted 11 female and 9 male healthy volunteers aged between 34 and 63 years (mean 41.7 ± 1.7), with no evidence of dry eye symptoms or signs.

Schirmer strips were used to measure tear secretion (Schirmer I test) and to collect tears. The measurements were made early in the morning, before the instillation of any ocular medications. Every patient used artificial tears (Viscofresh 1%; Allergan, Irvine, CA) at least five times per day. In addition, both groups were invited to complete the Dry Eye Questionnaire (DEQ) developed by Begley et al.13

Trials

Tear Collection. Tear secretion was measured by using the Schirmer I test (without anesthesia). The tear collection was always performed according to Van Bijsterveld criteria.14 The Schirmer strip was placed on the temporal tarsal conjunctiva of the lower lid for 5 minutes with the eyes closed. The volume of tears, in millimeters of moistened strip, was recorded, and the Schirmer strips were placed in tubes (Eppendorf, Fremont, CA) containing 500 μL of purified water (Ultrapure; Millipore, Billerica, MA). The samples were then frozen until they were analyzed by high pressure liquid chromatography (HPLC).

Tear Processing and HPLC Analysis. After thawing, the samples were strongly vortexed for 5 minutes. The strips were carefully rinsed, and the liquid in the tube was heated in a 100°C bath for 20 minutes, to precipitate the proteins. To pellet the proteins, we centrifuged the tubes at 4000 rpm for 30 minutes. Diadenosine polyphosphates are not degraded by this treatment, as previously demonstrated.15 Supernatants were chromatographed through a silica-based hydrophilic anion exchanger (SEP-PAK Accell QMA cartridge; Waters, Milford, MA). Briefly, 250 μL of the supernatant was passed through the cartridges which had been equilibrated with 3 mL of distilled water. Elution of the nucleotides and dinucleotides was performed by applying 1 mL of a solution containing 0.2 M KCl and 0.1 M HCl. Before injection into the HPLC, the samples were neutralized with KOH. The eluents were injected at a volume of 10 to 100 μL.

Determination and quantification of diadenosine polyphosphates were performed by HPLC. The chromatographic system consisted of an isocratic HPLC pump (model 1515; Waters), a dual-absorbance detector (model 2487; Waters), and an injector (Reodyne; Perkin Elmer, Boston, MA), all managed by the software (Breeze; Waters). The system was equilibrated overnight with the following mobile phase: 0.1 M KH2PO4, 2 mM tetrabutyl ammonium, and 17% acetonitrile (pH 7.5).17 Detection was monitored at 260 nm wavelength. All the peaks identified as putative dinucleotides were taken for phosphodiesterase treatment. Phosphodiesterase from Crotalus adamanteus (EC 3.1.4.1) from Sigma-Aldrich (St. Louis, MO), at a concentration of 1.0 U/mL, was incubated at room temperature for 30 minutes with the corresponding putative dinucleotide. The digestion products...
were analyzed by HPLC. Peaks were transformed into concentrations by means of external standards of known concentrations of diadenosine polyphosphates.

In the tear samples, two peaks were identified as Ap₄A and Ap₅A, according to the retention times (Fig. 1A). To confirm the nature of the putative Ap₄A and Ap₅A, both peaks were individually collected and treated with phosphodiesterase from Crotalus adamanteus (Figs. 1B, 1C). The putative Ap₄A produced two peaks after the enzyme action that were identified as AMP and ATP, whereas the putative Ap₅A produced AMP and adenosine tetraphosphate. These results indicate that these two original peaks corresponded to Ap₄A and Ap₅A. The comparison of the peaks with the corresponding standards permitted quantification of these two dinucleotides in the samples (Fig. 1D).

Dry Eye Questionnaire (DEQ)
This questionnaire was designed to assess the prevalence, frequency, and diurnal severity of ocular surface symptoms such as discomfort, dryness, visual changes, soreness, and irritation, grittiness and scratchiness, foreign body sensation, burning, light sensitivity, and itching. In the first part of the questionnaire, inquiries regarding the frequency of eight typical symptoms of dry eye were made, in which the patients chose between four different frequencies (rarely, sometimes, frequently, or constantly). The last part of the questionnaire includes questions about computer use, allergies, use of systemic and ocular medications, and ambient conditions that cause dry eye symptoms.13

The validation of the Spanish translation of the DEQ was supervised by the Psychology Faculty of the Universidad Complutense de Madrid and the original authors, Carolyn Begley and Robin Chalmers, who verified with a Spanish translator the concordance between the English and Spanish documents.

### Table 1. Concentrations of Nucleotides, Ap₄A and Ap₅A, in Control Group and Different Dry Eye Group Severity

<table>
<thead>
<tr>
<th></th>
<th>Control Group (n = 20)</th>
<th>SS Subgroup (n = 8)</th>
<th>Non-SS Group (n = 12)</th>
<th>SS Subgroup (n = 4)</th>
<th>Non-SS Subgroup (n = 34)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ap₄A</td>
<td>0.126 (0.027)</td>
<td>4.09 (1.36)</td>
<td>10.68 (0.54)</td>
<td>0.47 (0.20)</td>
<td>0.587 (0.058)</td>
</tr>
<tr>
<td>Ap₅A</td>
<td>0.044 (0.023)</td>
<td>39.51 (8.45)</td>
<td>12.45 (0.88)</td>
<td>8.3 (3.27)</td>
<td>0.058 (0.003)</td>
</tr>
</tbody>
</table>

The data presented are mean micromolar concentrations of the two dinucleotides (SD). Groups have been divided as normal or low tear production SS patients and compared with dry eye non-SS patients who have normal or low tear production.9

### Statistical Analysis

The values presented are the mean ± SEM of the experiments performed (version 15.0 for Windows; SPSS, Inc., Chicago, IL). In the case of Sjögren syndrome subgroups values also are presented as the median (first quartile; third quartile). Normal distribution of variables was assessed by the Kolmogorov-Smirnov normality test.

Sample size calculations were performed with statistical software (Granmo 6.0; Institut Municipal d’Investigació Mèdica, Barcelona, Spain). With an accepted two-sided statistical significance threshold of 0.05 and a β risk of 0.20 and taking into account a 2:1 group ratio (control group to Sjögren syndrome group), 16 subjects were needed in first group and 8 in the second, to find statistically significant differences.

Differences between the Sjögren syndrome and control groups in diadenosine polyphosphates levels were estimated by the Student’s t-test for independent samples. Because sample sizes of these groups are small, Mann-Whitney U tests were used to compare values between the Sjögren subgroups.

Correlations between ordinal data in the Sjögren syndrome group were assessed by Pearson bivariate regression, and one-way analysis of variance (ANOVA) was used to assess the correlation between diadenosine polyphosphate concentrations and each symptom’s frequency.

For statistical analysis of DEQ symptoms data, we used the authors’ test criteria.15 These criteria indicate that patients who responded frequently or constantly to the frequency questions were symptomatic. We regarded responses with a score of 4 or 5 to the questions about intensity as signifying that the patient was experiencing intense symptoms. The χ² test was used to contrast frequencies between groups, and the Wilcoxon test was used to compare the diurnal severity and diurnal (first quartile; third quartile). Normal distribution of variables was assessed by the Kolmogorov-Smirnov normality test.

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**FIGURE 2.** Schirmer I test in the Sjögren syndrome patients and normal subjects. Tear secretion was significantly lower in the Sjögren syndrome group than in the control group. The Sjögren syndrome group could be split into two subgroups, one with low tear secretion and one with normal tear secretion. *P < 0.05, Student’s t-test.
changes in symptom intensities (morning to evening). $P < 0.05$ was statistically significant.

**RESULTS**

**Tear Volume Results**

The tear volumes collected in the Sjögren syndrome group resulted in a mean Schirmer strip wetting result of $6.2 \pm 2.1$ mm ($n = 12$). Four Sjögren patients had a Schirmer test of more than 5 mm (Sjögren syndrome normal tear subgroup) and a mean of $12.3 \pm 1.2$ and median of $11$ mm (10; 13), and eight patients had results below 5 mm (Sjögren syndrome low tear volume subgroup), with a mean of $1.0 \pm 0.3$ mm and median of 0.75 mm (first and third quartiles: 0.50; 1.00; $P < 0.001$; Sjögren syndrome low tear subgroup versus Sjögren syndrome normal tear subgroup; Mann-Whitney U test). The mean tear volume in the control group was $19.9 \pm 2.3$ mm ($n = 20$; $P < 0.001$ control group versus Sjögren syndrome group and control group versus Sjögren syndrome normal tear subgroup; Student’s t-test; Fig. 2).

**Nucleotide Analysis in Tears**

The mean concentrations of Ap$_4$A, and Ap$_5$A in the Sjögren syndrome group were $2.54 \pm 1.02$ and $26.13 \pm 6.95$ μM, respectively. In the subgroup of Sjögren syndrome with normal tear secretion ($n = 4$), the concentrations of Ap$_4$A and Ap$_5$A were $0.47 \pm 0.20$ μM (median, 0.41 μM; first and third quartiles, 0.12; 0.86) and $8.3 \pm 3.27$ μM (median, 6.4 μM; 3.01; 14.60), respectively. The subgroup of Sjögren syndrome with low tear secretion ($n = 8$) showed dinucleotide concentrations of $4.09 \pm 1.36$ μM (median, 4.26 μM; 0.23; 6.5) and $39.51 \pm 8.45$ μM (median 36.6 μM; 17.85; 59.14) for Ap$_4$A and Ap$_5$A, respectively. These values were significantly different between the subgroups ($P < 0.05$ and $P < 0.02$ for Ap$_4$A and Ap$_5$A, respectively; Mann-Whitney U test). In the control group the concentrations of Ap$_4$A and Ap$_5$A were $0.13 \pm 0.03$ and $0.04 \pm 0.02$ μM, respectively (P $< 0.05$ to Ap$_4$A and $P < 0.01$ to Ap$_5$A; control group versus Sjögren syndrome group; Student’s t-test; Table 1, Fig. 3).

**DEQ Results**

In the Sjögren syndrome group, the most common responses to all the symptoms were frequently or constantly. In patients with Sjögren syndrome who had a low tear volume, the most frequent symptoms were eye discomfort, dry eyes, irritated eyes, and sensitivity to light. In patients who had a normal tear volume, all had symptoms of foreign body sensation, irritation, and sensitivity to light. The results of the

<table>
<thead>
<tr>
<th>Symptom</th>
<th>Control Group ($n = 20$)</th>
<th>SS Group ($n = 20$)</th>
<th>Low Tear Subgroup ($n = 8$)</th>
<th>Normal Tear Subgroup ($n = 4$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Discomfort</td>
<td>3 (15)</td>
<td>12 (100)</td>
<td>8 (100)</td>
<td>4 (100)</td>
</tr>
<tr>
<td>Dryness</td>
<td>1 (5)</td>
<td>9 (92)</td>
<td>8 (100)</td>
<td>3 (75)</td>
</tr>
<tr>
<td>Grittiness</td>
<td>0 (0)</td>
<td>6 (50)</td>
<td>5 (63)</td>
<td>1 (25)</td>
</tr>
<tr>
<td>Burning</td>
<td>0 (0)</td>
<td>5 (42)</td>
<td>4 (50)</td>
<td>1 (25)</td>
</tr>
<tr>
<td>Itching</td>
<td>1 (5)</td>
<td>7 (67)</td>
<td>5 (63)</td>
<td>3 (75)</td>
</tr>
<tr>
<td>Foreign body</td>
<td>0 (0)</td>
<td>8 (75)</td>
<td>5 (63)</td>
<td>4 (100)</td>
</tr>
<tr>
<td>Irritation</td>
<td>0 (0)</td>
<td>11 (92)</td>
<td>7 (88)</td>
<td>4 (100)</td>
</tr>
<tr>
<td>Light sensitivity</td>
<td>4 (20)</td>
<td>12 (100)</td>
<td>8 (100)</td>
<td>4 (100)</td>
</tr>
<tr>
<td>Blurring</td>
<td>1 (5)</td>
<td>6 (50)</td>
<td>5 (63)</td>
<td>1 (25)</td>
</tr>
</tbody>
</table>

Symptom distribution and frequency according to patients’ reports. The data are the number of patients reporting the respective symptom and, in parentheses, the percentage of patients reporting that symptom.
control group are shown in Table 2. Significant differences between the Sjögren syndrome groups and control group were observed ($\chi^2$ test, $P < 0.01$).

Patients assessed the intensity of the symptoms on a scale of 1 to 5, with 1 being the lowest and 5 the highest intensity, and we deemed a symptom intense when patients reported 4 or 5, according to the DEQ authors’ criteria. In patients with Sjögren syndrome, symptoms such as discomfort, dryness, foreign body, irritated eyes, and blurred vision increased in intensity in the evening. The increase in patients with intense symptoms was significantly greater in the low tear subgroup than in the normal tear subgroup (Fig. 4). The increase in the percentage of patients at the end of the day in the subgroup of Sjögren syndrome patients with low tears was statistically significant (Wilcoxon test, $P < 0.05$) for four of eight symptoms studied. In contrast, in the normal tears subgroup, there were no significant differences for any symptom (Table 3). There were no daytime differences in

![Figure 4](http://iovs.arvojournals.org/pdfaccess.ashx?url=/data/Journals/IOVS/932968/ on 10/20/2016)
the percentage of patients with intense symptoms for the control group.

**Correlations**

In the Sjögren syndrome group, the Pearson’s correlation coefficient was $-0.59$ ($P = 0.002$) for tear volume and $A_{p4A}$. The value of this coefficient indicates a low to moderate degree of correlation and the negative sign means that the lower the result of the Schirmer test, the greater the level of $A_{p4A}$. With respect to nucleotides, the Pearson’s correlation coefficient showed a moderate to high degree of correlation for $A_{p4A}$ ($P = 0.024$) with $A_{p5A}$ in the Sjögren syndrome group (Table 4).

In a previous work, it was possible to demonstrate the existence of differences between $A_{p4A}$ and $A_{p5A}$ concentrations and symptomatic and asymptomatic dry eye patients. In the present study, when trying to determine whether there were differences between any reported symptom and the concentrations of these dinucleotides in Sjögren syndrome patients, we did not find a statistically significant correlation between any reported symptom and the concentrations of these dinucleotides in Sjögren syndrome patients by ANOVA ($P > 0.05$).

**DISCUSSION**

In the present work, the concentrations of $A_{p4A}$ and $A_{p5A}$ in patients with Sjögren syndrome were investigated. These values were increased 42- and 595-fold for $A_{p4A}$ and $A_{p5A}$, respectively, compared with the control group. In the Sjögren syndrome subgroup with low tear secretion, the results showed that these patients had higher levels of the adenine dinucleotides $A_{p4A}$ and $A_{p5A}$ in their tears than did Sjögren syndrome patients with normal tear secretion. These facts could suggest that these dinucleotides are potential tools for the diagnosis of dry eye, as previously related.9

If we compare the data of Sjögren syndrome patients with those of non-Sjögren dry eye symptomatic subjects presented in a previous work (Table 1), the concentration of $A_{p4A}$ in patients with Sjögren syndrome with low tear secretion was less than half that in those non-Sjögren dry eye symptomatic patients with reduced tear secretion. In the Sjögren and the non-Sjögren dry eye groups with normal tear secretion, there were no differences in the concentrations of $A_{p4A}$. However, $A_{p5A}$ appeared in higher concentrations in patients with Sjögren syndrome compared with non-Sjögren dry eye subjects. This fact could indicate that $A_{p4A}$ can be a differentiating factor between Sjögren syndrome and non-Sjögren dry eye.

Several works have reported that the ocular surface in Sjögren syndrome patients is altered and damaged. A positive ocular staining with fluorescein and rose bengal has been described in these subjects.18,19 The diadenosine polyphosphates have a hastening effect on the corneal re-epithelialization, and a greater release of $A_{p4A}$ and $A_{p5A}$ in tears will therefore help naturally to facilitate the rate of wound healing.11 It has been reported that an increased blinking frequency enhances the levels of nucleotides in tears, because of the shear stress provoked by the eyelids on the ocular surface, and an increase in the blinking frequency of dry eye patients has been reported.20 The damage to the ocular surface and the increased blinking rate in Sjögren syndrome patients could explain why the presence of diadenosine polyphosphates in these patients is higher than in healthy individuals.

**Table 3. Results of Wilcoxon Signed-Rank Test Comparing the Diurnal Intensity of Symptoms**

<table>
<thead>
<tr>
<th>Symptom</th>
<th>Control Group (n = 20)</th>
<th>SS Group (n = 12)</th>
<th>SS Low Tear Subgroup (n = 8)</th>
<th>SS Normal Tear Subgroup (n = 4)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Discomfort</td>
<td>0.56</td>
<td>0.04*</td>
<td>0.01†</td>
<td>0.56</td>
</tr>
<tr>
<td>Dryness</td>
<td>0.66</td>
<td>0.31</td>
<td>0.02*</td>
<td>0.79</td>
</tr>
<tr>
<td>Grittiness</td>
<td>—‡</td>
<td>0.31</td>
<td>0.15</td>
<td>—‡</td>
</tr>
<tr>
<td>Burning</td>
<td>—‡</td>
<td>0.31</td>
<td>0.08</td>
<td>—‡</td>
</tr>
<tr>
<td>Itching</td>
<td>0.56</td>
<td>0.31</td>
<td>0.08</td>
<td>0.65</td>
</tr>
<tr>
<td>Foreign body</td>
<td>—‡</td>
<td>0.18</td>
<td>0.15</td>
<td>0.78</td>
</tr>
<tr>
<td>Irritation</td>
<td>—‡</td>
<td>0.04*</td>
<td>0.04†</td>
<td>—‡</td>
</tr>
<tr>
<td>Blurring</td>
<td>0.66</td>
<td>0.03*</td>
<td>0.01†</td>
<td>0.56</td>
</tr>
</tbody>
</table>

* $P < 0.05$.
† $P < 0.01$.
‡ Same intensity in the morning and evening in all patients.

**Table 4. Correlation among Diadenosine Polyphosphates, Schirmer Test and Age in Sjögren Syndrome Patients**

<table>
<thead>
<tr>
<th></th>
<th>$A_{p4A}$ Pearson correlation</th>
<th>$A_{p5A}$ Pearson correlation</th>
<th>Schirmer Test Pearson correlation</th>
<th>Age Pearson correlation</th>
</tr>
</thead>
<tbody>
<tr>
<td>$A_{p4A}$</td>
<td>1</td>
<td>$-0.76$</td>
<td>$-0.43$</td>
<td>0.12</td>
</tr>
<tr>
<td>$A_{p5A}$</td>
<td>$-0.76$</td>
<td>1</td>
<td>$-0.59$</td>
<td>0.28</td>
</tr>
<tr>
<td>Schirmer test</td>
<td>$-0.43$</td>
<td>$-0.59$</td>
<td>1</td>
<td>$-0.20$</td>
</tr>
<tr>
<td>Age</td>
<td>0.11</td>
<td>0.02*</td>
<td>$-0.20$</td>
<td>0.41</td>
</tr>
</tbody>
</table>

Pearson bivariate regression test to compare the possible correlation among nucleotides, Schirmer test and age.

* $P < 0.01$ (two-tailed).
† $P < 0.05$ (two-tailed).
It has been shown that ApA and Ap₄A stimulate tear secretion, and it appears that they are naturally released and serve to increase the tear volume and quality by stimulating the P₂Y₂ receptors present in meibomian and accessory glands. Although the presence of P₂Y₂ receptors in the lacrimal gland has not yet been reported, these receptors have been found in the lacrimal gland of New Zealand White rabbit (Hoyle CHV, personal communication, 2009). Their presence in the salivary glands has been demonstrated.

Schrader et al. indicated that P₂Y₂ receptor expression is upregulated in the submandibular gland of the NOD.B10 mouse, a model of Sjögren syndrome, to provide a pathway for fluid secretion. Furthermore, Baker et al. reported that P₂Y₂ receptor activation in salivary gland cells increases the epidermal growth factor receptor (EGFR)-dependent expression of vascular cell adhesion molecule (VCAM)-1 and the binding of lymphocytes.

A similar effect may occur in the lacrimal gland of Sjögren syndrome patients, where the expression of the P₂Y₂ receptor may be augmented to increase tear secretion, but in fact provokes the lymphocyte infiltration that may lead to inflammation associated with Sjögren syndrome. Although ApA and Ap₄A are present at high concentrations in Sjögren syndrome patients, it seems to be that they are not able to stimulate more lacrimation. Instead they may be increasing lymphocyte infiltration. Further studies are needed to confirm our hypothesis that P₂Y₂ upregulation really occurs in lacrimal glands of Sjögren syndrome patients and to establish the purpose of this upregulation.

In normal individuals dinucleoside polyphosphates increase tear production. In contrast, some Sjögren patients lack this dinucleotide stimulation. Presumably, Sjögren patients with normal tear production have the benefit of the secretagogue action triggered by these molecules, in clear contrast with those who do not seem to be sensitive to these dinucleotides. It could be the case that the difference between these two subpopulations of Sjögren patients is due to factors such as a poorer tear quality that makes it more evaporative, or some problems with the lachrymal apparatus. There is no evidence of dysfunction of the P₂Y₂ receptors in the lachrymal gland, but such dysfunction may explain the lack of secretagogue activity for these substances in some Sjögren patients. This point should be investigated more thoroughly.

Concerning the Schirmer test results, it may be surprising that in a disease whose main sign is aqueous tear deficiency, patients have values of tear secretion above 10 mm. Others studies have found similar results for the Schirmer test. This is probably due to the different severities of the disease and the poor sensitivity of the test.

Another interesting point is the lack of correspondence between the increase in tear volume and the dinucleotide concentration. The present experiments demonstrated that when there was an increase in tear production, there was not a linear change in the dinucleotide concentration. Indeed dinucleotides were not “diluted” as expected, perhaps because of the way these compounds are released. In particular, diadenosine polyphosphates are released as a consequence of mechanical stress, which very often relates to an increase in the blinking frequency due to discomfort.

Discomfort, light sensitivity, and dryness were the most frequent and intense symptoms, which is consistent with the results reported by other researchers. Sjögren syndrome patients reported that their symptoms increased later in the day, and they were more pronounced in the low tear secretion group. Versura et al. suggested an environment or task-related etiology for dry eye symptoms as the main factor in increased intensity at the end of the day.

There were no statistically significant differences in ApA and Ap₄A concentrations in tears from normal humans aged from 20 to 50 years (our unpublished data, 2010). Therefore, the difference in ages of the control and Sjögren syndrome groups is not a factor in the difference in diadenosine polyphosphate concentrations.

In conclusion, concentrations of diadenosine polyphosphates are increased in tears of Sjögren syndrome patients, especially Ap₄A, which could be used as a diagnostic tool for this condition. In addition, we conclude that ApA is increased in pathologic dry eye with respect to healthy eyes, confirming its role as an objective marker of dry eye.

Acknowledgments

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