

1 **Optimization of the Fenton treatment of 1,4-dioxane and on-line**
2 **FTIR monitoring of the reaction**

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4 *Noemí Merayo, Daphne Hermosilla* , Luis Cortijo and Ángeles Blanco*

5
6 Department of Chemical Engineering, Complutense University of Madrid, Facultad de Ciencias
7 Químicas, Ciudad Universitaria s/n, 28040 Madrid, Spain.

8
9 * Corresponding author: *Tel.: +34 91 394 4645; fax: +34 91 394 4243.*

10 E-mail address: dhermosilla@quim.ucm.es (D. Hermosilla).

11
12 E-mail address of each author in order of appearance:

13 nmerayoc@quim.ucm.es (N. Merayo)

14 dhermosilla@quim.ucm.es (D. Hermosilla)

15 lcortijo@quim.ucm.es (L. Cortijo)

16 ablanco@quim.ucm.es (A. Blanco)

17

1 ABSTRACT

2 1,4-dioxane is a non-biodegradable, toxic, hazardous, and priority pollutant widely used in the
3 chemical industry as a solvent; as well as it is a resulting by-product of many industrial
4 processes. The optimization of the Fenton treatment of 1,4-dioxane, and the on-line FTIR
5 monitoring of its degradation route, including the assessment of the enhancement of the
6 biodegradability of the solution along treatment are herein addressed. Besides the full removal of
7 1,4-dioxane, an 80% reduction of the chemical oxygen demand (COD) was achieved at the best
8 tested treatment conditions. Whether the used concentration of H₂O₂ was expectedly addressed
9 as the reaction factor most influencing the achieved COD removal at the end of the process; the
10 performance of the treatment under acid pH conditions showed to have just a slight influence,
11 thus supporting this process may suitably be performed at neutral pH value. On-line FTIR
12 monitoring of the process novelly provided the degradation route of 1,4-dioxane along its
13 oxidation treatment, as well as a comprehensive optimization of the Fenton process based on the
14 increase of the biodegradability of the solution and the reduction of the consumption of reagents.

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16 KEYWORDS: Advanced oxidation processes; Fenton method; FTIR; on-line reaction
17 monitorization; 1,4-dioxane oxidation.

18

1 **1. Introduction**

2 1,4-dioxane is an organic compound used as solvent in many industrial processes, as well as it is
3 generated as a by-product in several industrial chemical reactions [1]. The main environmental
4 concern regarding the potential release of this compound to the environment has recently
5 increased due to its major negative effects on health. Correspondingly, 1,4-dioxane is classified
6 as a priority pollutant by the US EPA [2]. Furthermore, it is a bio-recalcitrant and persistent
7 organic molecule that cannot be treated by conventional bio-treatment technologies [1,3]. In fact,
8 physical treatments, such as adsorption on activated carbon or air stripping, have not been
9 successful removing dioxane from water because of this inherent high aqueous solubility and its
10 low vapor pressure [3,4]. Moreover, it has been addressed that the chemical oxidation of 1,4-
11 dioxane by chlorine could lead to the formation of even more toxic compounds [1]. In
12 conclusion, traditional treatments have not been shown as effective degrading this substance.
13 Only distillation technology has efficiently been proved to separate 1,4-dioxane from the
14 solution due to its high volatility; but its use has a very high associated cost, particularly at very
15 low concentration levels [1,3,4].

16 Advanced oxidation processes (AOPs) may represent a feasible treatment alternative for
17 1,4-dioxane [4] because they can be applied at a lower economical cost just transforming this
18 persistent compound into simpler more biodegradable compounds that might be further degraded
19 by traditional biological treatment [5]. Particularly, Fenton method is one of the most commonly
20 used AOPs. It is based on the electron transfer between hydrogen peroxide and ferrous ion,
21 which acts as a homogenous catalyst, yielding hydroxyl radicals ($\text{OH}\cdot$) that are able to degrade a
22 wide range of organic compounds [6]. In general, the Fenton process is usually very efficient,
23 and it even implies a lower economical cost than the application of others AOPs [7]. Regardless
24 a high chemical oxygen demand (COD) removal efficiency could be achieved and a relative

1 overall environmental friendliness, a Fenton treatment of wastewater is limited by the final
2 production of iron sludge, which requires ultimate disposal [8], and the optimal pH range of
3 application, which is usually set at acid values ($\text{pH} \approx 3$) [9,10].

4 There are very few limited studies considering the treatment of 1,4-dioxane by Fenton's
5 reagent [1,3,11]. Although all these essays have assessed the improved elimination of 1,4-
6 dioxane and COD from wastewater, an increased biodegradability of the final solution, and the
7 reduced consumption of reagents; a comprehensive process optimization has not been addressed
8 yet. In fact, the identification of which by-products are formed along the degradation of 1,4-
9 dioxane by Fenton has not been addressed to date. Some authors have previously reported the
10 removal of 1,4-dioxane and other by-products that are formed along its treatment, but the
11 production of other chemical species along the reaction has not really been assessed in full yet
12 [1,11,12]. Only Stefan and Bolton [13] have previously proposed a degradation mechanism for
13 1,4-dioxane treating this chemical by UV/H₂O₂ and taking periodical samples that were analyzed
14 by gas chromatography (GC), solid phase microextraction (SPME), gas chromatography/mass
15 spectrometry (GC/MS), ion chromatography (IC), and high-performance liquid chromatography
16 (HPLC), in order to identify intermediate chemical species and by-products. The advantage of
17 monitoring similar processes using Fourier transform infrared spectroscopy (FTIR) has
18 previously been addressed identifying the compounds that are appearing and disappearing in the
19 solution along the reaction sampling aliquots at preselected time intervals [14, 15].

20 Therefore, the main objectives of this research are: (1) optimizing conventional Fenton
21 method as a promising alternative for the treatment of 1,4-dioxane; and (2) developing an FTIR-
22 based methodology enabling the on-line control of the use of reagents, and the production of
23 intermediates and by-product along the degradation of 1,4-dioxane.

24

1 **2. Material and methods**

2 *2.1. Materials and analytical methods*

3 All used chemicals were of analytical grade and supplied by PANREAC S.A. (Barcelona, Spain)
4 or Sigma-Aldrich (Highland, USA). The solutions were prepared in deionized water and kept in
5 the dark until use. All the analyses were made according to the standard methods for the
6 examination of water and wastewaters [16]. COD was measured by the colorimetric method at
7 600 nm using an Aquamate-spectrophotometer (Thermo Scientific AQA 091801, Waltham,
8 USA). Hydrogen peroxide concentration was analyzed by the titanium sulphate
9 spectrophotometric method [17].

10 In order to confirm FTIR results, 1,4-dioxane and ethylene glycol were identified and
11 quantified along the reaction by gas-liquid chromatography (GLC) using a 7980A instrument
12 (Agilent Technologies Inc., Palo Alto, CA) equipped with a flame ionization detector. Injector
13 and detector were respectively set up at 310 and 280 °C. Samples (2 µL) were injected using the
14 *pulsed-split* mode (*split* ratio 5:1) and analyzed in a TRB-FFAP (Teknokroma, Sant Cugat del
15 Vallès, Spain) fused silica column (30 m x 0.25 mm internal diameter x 0.25 µm film thickness),
16 with He (43 psi) as carrier gas, and the following temperature program: 80 °C to 240 °C at a 15
17 °C·min⁻¹ ramp rate after a 9 min initial hold. Peaks were identified on the basis of sample
18 coincidence to relative retention times of commercial standards. Quantification was performed
19 according to peak areas that were corrected by response factors that were calculated for each
20 compound using 1-butanol (250 mg·L⁻¹) as internal standard, and *GC-ChemStation Rev.B.04.02*
21 (*96*) software from Agilent.

22 Formic, oxalic, acetic, glycolic, and methoxyacetic acids were also complementarily
23 identified and quantified by ion chromatography (IC) using a Dionex DX-500 device (Thermo
24 Scientific, Sunnyvale, CA) equipped with a conductivity detector. A 40 to 60 mM NaOH

1 gradient was used as the eluent for measurement keeping the flow at $1.5 \text{ L} \cdot \text{min}^{-1}$. The injection
2 loop was $75 \mu\text{L}$. An AS11HC Ion Pac ionic resin column was used aided with a previous Anion
3 Trap Column (ATC3) and a AG11-HC guard column. Peaks were identified and quantified on
4 the basis of sample coincidence to relative retention times and standard concentrations of
5 commercial standards.

6

7 *2.2. FTIR analytical equipment*

8 Mettler-Toledo ReactIR iC10 (Columbia, USA) is a Fourier transform infrared (FTIR)
9 spectrometer that measures chemical species as they react over a period of time. It uses a
10 Mercury Cadmium Telluride (MCT) detector that is cooled by liquid nitrogen. Measurements are
11 optically taken using a diamond tipped probe with a 1 meter fiber optic conduit. The system
12 should be purged (using instrument grade air, nitrogen or other suitable inert gas) in order to
13 prevent water vapour from collecting inside the optics, which might otherwise obscure spectral
14 data.

15 Data acquisition in the absorbance scale was taken from 2000 to 750 cm^{-1} with an 8 cm^{-1}
16 nominal resolution; and 256 scans were co-added for each spectrum. A background on pure
17 water was carried out just before performing each spectral record under the same resolution and
18 scanning conditions that were used for the trials. Real-time component analyses were run using
19 ConciRT software (Mettler-Toledo, Columbia, USA), which calculates associated component
20 spectra, and relative concentration profiles.

21

22 *2.3. Experimental procedure*

23 Experiments were performed in a 500 mL glass reactor placed on a magnetic mixing device.
24 Temperature was adjusted to the desired value ($25 \text{ }^\circ\text{C}$) using a water heater and circulator. pH

1 was monitored along the process, and adjusted to the desired value (± 0.1) using $1 \text{ mol}\cdot\text{L}^{-1}$
2 sodium hydroxide, or $1 \text{ mol}\cdot\text{L}^{-1}$ sulphuric acid, as required. Considering 1,4-dioxane is degraded
3 to ethylene glycol at a very slow rate at acid pH values, whereas it is ionized and keeps a stable
4 structure under basic ones, pH adjustment was carried out after adding dioxane ($247.8 \text{ mg}\cdot\text{L}^{-1}$;
5 initial COD= $450 \text{ mgO}_2\cdot\text{L}^{-1}$) when performing trials under acid conditions, and before its
6 incorporation to the solution otherwise. After temperature and initial pH adjustment, ferrous
7 sulphate was added to the solution at the ferrous ion concentration specified in the experimental
8 design described next to optimize treatment results. Hydrogen peroxide (30% w/v) was then
9 added in batch mode until the designed concentration was also reached. Treatments were run
10 until all the added H_2O_2 was totally consumed, so maximum COD removal was achieved for a
11 set of reaction variables values.

12 Aliquots of the treated solution were periodically sampled with a syringe along the trials.
13 These samples were adjusted to $\text{pH}\approx 9.0$ adding 40% sodium hydroxide (NaOH), and then
14 centrifuged during 15 min at 2000 rpm. COD and H_2O_2 concentrations were measured in the
15 collected supernatant. H_2O_2 concentration values were used to correct COD values according to
16 Hermosilla et al. [18].

17 Initial reaction conditions and concentrations of reagents chosen to perform on-line FTIR
18 control experiments were determined after optimizing the Fenton treatment of 1,4-dioxane. In
19 general, the experimental procedure used for FTIR trials was the same, despite the concentration
20 of 1,4-dioxane was $0.07 \text{ mol}\cdot\text{L}^{-1}$, and the addition of hydrogen peroxide was carried out in
21 continuous mode ($8.8 \text{ mmol H}_2\text{O}_2\cdot\text{min}^{-1}$ to a total 100 mL of reaction solution) because it allows
22 to understand better the reaction process; besides it has previously shown better results than
23 batch mode in this particular process [18, 19].

1 The main compounds that actual scientific literature proposes to be considered in the
2 degradation route of 1,4-dioxane by oxidation are: ethylene glycol diformate, glycolic acid,
3 glyoxylic acid, methoxyacetic acid, formic acid, acetic acid and oxalic acid [1,3,11,13, 20]. Their
4 reference spectra were first experimentally collected by FTIR (Figure 1). Despite the likeness of
5 these compounds, the proper interpretation of mixture spectra (recorded during Fenton
6 processes) have been successfully carried out with the help of the mathematical algorithms
7 integrated in ConcIRT software, and comparing spectra of pure solutions with those obtained
8 along the process. Water spectra were subtracted from the obtained on-line spectra in all
9 experimental runs.

11 *2.4. Experimental design*

12 Response surface methodology (RSM) was used to optimize the Fenton treatment of 1,4-dioxane
13 statistically assessing the significance of the effects of the considered individual factors, and the
14 interactions between them, that influence the process [21, 22]. In addition, Pareto charting was
15 used to highlight the most important factors that influenced the treatment efficiency. In short,
16 this methodology allows determining optimum operation conditions for the system, as well as
17 the region in which the operating specifications are satisfied [22]. Experimental design,
18 regressions, response surfaces, and Pareto charts, were run by Systat 13 software (SYSTAT
19 Software Inc., Chicago, USA).

20 pH, temperature, the concentrations of reagents, and reaction time, are the main factors
21 that may influence the Fenton treatment of an organic compound. Among them, temperature has
22 previously been reported not to affect much between 25 to 45 °C, and reaction time is really
23 dependent on the concentrations of reagents [18]. Therefore, experiments were designed to
24 optimize pH, hydrogen peroxide and ferrous ion concentrations. Based upon previously reported

1 results for optimizing Fenton treatment of organics [23], RSM was actually performed
2 considering pH values of 2.8, 5.0 and 7.2; and H₂O₂ concentration dependent ratios of
3 [H₂O₂]₀/COD₀ = 1.063, 2.125 and 4.250; and [H₂O₂]₀/[Fe²⁺]₀ molar ratios of 1.0, 5.0 and 10.0.

4 In short, RSM regression analysis was performed according to a full factorial design
5 (N=27) using the following quadratic model:

$$6 \quad Y = b_0 + \sum_{i=1}^k b_i X_i + \sum_{i=1}^k b_{ii} X_i^2 + \sum_{i=1}^k \sum_{j=1}^k b_{ij} X_i X_j$$

7 Where Y (COD removal) is the process response dependent variable; X_i are the
8 considered independent variables (X₁ = [H₂O₂]₀/COD₀; X₂ = [H₂O₂]₀/[Fe²⁺]₀; X₃ = pH); and b_i,
9 b_{ii} and b_{ij} are the corresponding regression coefficients.

10

11 **3. Results and discussion**

12 *3.1. Optimization of the conventional Fenton treatment of 1,4-dioxane*

13 pH is one of the most important factors influencing the final result of a Fenton process because it
14 mainly controls the speciation of iron and the stability of hydrogen peroxide [24, 25].

15 In short, iron precipitation as Fe(OH)₃ is avoided under acid conditions because the solubility of
16 iron increases [26] and, as a consequence, the efficiency of the process also increases. In
17 addition, hydrogen peroxide decomposition is also impeded under acid environments, favoring
18 its stabilization [27]; as well as the scavenger function of inorganic carbon is also prevented
19 because it should mainly have already been removed. Nonetheless, further significance may be
20 found in the case of the Fenton treatment of 1,4-dioxane because of the structural changes this
21 compound shows in relation to the pH value of the solution.

22 The resulting estimations of the regression coefficients of the quadratic model used to
23 optimize the Fenton treatment of 1,4-dioxane by RSM (Figure 2) are shown in Table 1. Whether

1 a positive value of one of these coefficients indicates a positive relation of the associated factor
2 to COD reduction; negative coefficients mean a lower COD removal would be achieved at a
3 higher value the related factor. In particular, the negative coefficient related to the pH factor
4 means that higher COD reductions were achieved working at lower pH values (Figure 2); as well
5 as higher doses of H₂O₂ (a higher [H₂O₂]₀/COD₀ ratio, actually) increased the difference between
6 the reduction of the COD that was achieved working at those designed lower and higher pH
7 values. In fact, the effect of pH was slight at the designed lower dose value of H₂O₂ (less than
8 10%).

9 The optimum pH value for performing the Fenton treatment of 1,4-dioxane resulted 2.8,
10 as it has previously been also reported, although the effect of pH was not as relevant as it has
11 been addressed before [18, 28]. The highest influence of pH was exerted at a ratio of
12 [H₂O₂]₀/COD₀=4.250, obtaining a good COD removal result (≈55%) when the process was
13 performed at pH=7.2 if it is compared to the optimized result at pH = 2.8 (COD removal≈65%;
14 Figure 2). This shorter difference in the efficiency of the process being performed at both pH
15 values was probably the result of the structural changes that are promoted in the molecule of 1,4-
16 dioxane when the pH value changes from acid to basic. In fact, the ionization of 1,4-dioxane
17 under basic reaction conditions favors its reaction with hydroxyl radicals. As a result, the use of
18 chemicals for controlling the pH may be partially counterbalanced as the main drawback for the
19 implementation of Fenton process at industrial scale [29].

20 In general, the optimization of chemical reagents use is an important issue for the
21 industrial application of Fenton processes due to its related high economical cost. A higher COD
22 removal was achieved when a higher ratio of [H₂O₂] was used in relation to the initial COD
23 value of the solution (Figure 2) because as more hydrogen peroxide is available, the generation
24 of hydroxyl radical is also higher; although an excessive amount of H₂O₂ may scavenge

1 hydroxyl radicals reducing the efficiency of the treatment [18]. In fact, this ratio between the
2 used H_2O_2 concentration and the initial value of the COD in the solution was addressed to show
3 the highest influence on the reduction of the COD (Figure 3). As a consequence, COD removal
4 losses derived from not performing the process under such a low optimum pH value did not
5 result so important when the optimum amount of H_2O_2 is added to the solution (Figure 2). In
6 addition, the molar rate between reagents ($[\text{H}_2\text{O}_2]_0/[\text{Fe}^{2+}]_0$) resulted the second most important
7 parameter influencing process efficiency. Its associated negative coefficient in the resulting RSM
8 model (Table 1) implies that COD removal was enhanced when this ratio between reagents was
9 lower. Therefore, considering certain amount of H_2O_2 is added to perform treatment, the higher
10 tested related supply of Fe^{2+} yielded the higher removal of COD.

11 In summary, the best tested reaction conditions achieved a 65% COD removal ($\text{pH} = 2.8$,
12 $[\text{H}_2\text{O}_2]_0/\text{COD}_0 = 4.250$ and $[\text{H}_2\text{O}_2]_0/[\text{Fe}^{2+}]_0 = 1$). The COD remaining in the solution after this
13 optimum treatment was characterized by the high content of volatile fatty acids ($72.5 \text{ mg}\cdot\text{L}^{-1}$),
14 which represents a high increase of biodegradability in the final solution. Therefore, the
15 oxidative degradation of 1,4-dioxane at least resulted in the final persistence of more
16 biodegradable organic acids, which is in agreement with previous results reporting an almost
17 total destruction of 1,4-dioxane and a progressive decrease of the pH of the solution due to the
18 formation of organic acids [1]. In addition, a poor elimination of the DOC further supported the
19 degradation of 1,4-dioxane into more biodegradable products rather than to CO_2 , which is also in
20 agreement with previous related results [11].

21 In addition, more feasible industrial treatment conditions were chosen to perform further
22 trials exploring the potential biodegradability enhancement of wastewater containing dioxane,
23 namely: $\text{pH}=2.8$, $[\text{H}_2\text{O}_2]_0/\text{COD}_0=2.125$ and $[\text{H}_2\text{O}_2]_0/[\text{Fe}^{2+}]_0=5$. Higher doses of the reagents
24 involve a proportional increase in the costs of chemicals and disposing iron sludge. Therefore,

1 optimum reaction values resulting from RSM modeling are not really justified for real
2 applications. As a result, a 43% COD removal was achieved performing the Fenton treatment of
3 1,4-dioxane under these reaction conditions; as well as the final presence of volatile fatty acids
4 reached $58 \text{ mg}\cdot\text{L}^{-1}$, which also means a significant increase of the biodegradability of the final
5 solution.

6

7 *3.2. On-line FTIR monitoring of the degradation of 1,4-dioxane by Fenton oxidation*

8 Concentration profiles of the main compounds that were generated in the solution along the
9 Fenton oxidation of 1,4-dioxane (Figure 4) were obtained monitoring the process with an on-line
10 FTIR probe. The ConClRT software identified the compounds that were being on-line detected
11 by FTIR comparing them to the previously obtained reference FTIR spectra of these compounds
12 (Figure 1). Initially, FTIR spectra did not show any change after the addition of a low
13 concentration of H_2SO_4 at the studied wavenumber spectral region; as well as it was shown that
14 the degradation of 1,4-dioxane into ethylene glycol under acid reaction conditions resulted too
15 slow to be detected at the beginning of the reaction (Figure 5). In fact, the absence of ethylene
16 glycol at this point was also checked by chromatographic analysis before the addition of
17 Fenton's reagent, after which certain very early changes can be noticed in the spectra. The
18 dissolution of ferrous sulphate heptahydrate reported a peak at about 1100 cm^{-1} , which surely
19 represents sulphate ion [30]. The evolution of iron content cannot be followed separately.

20 1,4-dioxane and $\text{FeSO}_4\cdot 7\text{H}_2\text{O}$ were already present in the solution before any H_2O_2 was
21 added. A red-brown colour appeared just after hydrogen peroxide was incorporated, denoting
22 iron oxidation from pale-green ferrous ion to red-orange ferric one. A final intense orange color
23 remained in the solution during almost all the process. When 8.8 mmol of H_2O_2 were already
24 added to the reaction medium, ethylene glycol began to increase its presence, as well as glycolic

1 acid and oxalate anion; whereas 1,4-dioxane began to decrease (Figure 5). After 35 mmol of
2 H₂O₂ were fully added, glycolic acid and oxalate anion reached a maximum of absorbance and
3 began to decrease their presence thereafter. On the other hand, the presence of formic acid was
4 newly detected at this point, and increased much its content until reaching a constant value at the
5 end of the reaction. Thereafter, the reaction evolved until the complete reduction of 1,4-dioxane
6 just when all the H₂O₂ dosage (70.5 mmol) was added. In addition, the profile of ethylene glycol
7 showed a slightly decreasing tendency at the same time that glycolic acid and oxalate anion
8 increased their presence at a similar linear pace, denoting certain relationship between both
9 trends towards the end of the process.

10 Chromatographic analyses confirmed the presence of all these compounds at the same
11 moments of the reaction and the full removal of 1,4-dioxane at the end of the Fenton treatment.
12 Therefore, the same process evolution shown in Figure 5 was confirmed by the performed
13 chromatographic determinations at singular reaction moments. Particularly, IC analyses
14 indicated that, whether glycolic acid was produced in a small quantity, the presence of oxalate
15 was very high. This might have been the result of a fast degradation of glycolic acid into oxalic
16 acid, which may actually be the main responsible of the detected trend.

17 The pH-dependence of oxalic acid caused the appearance of the corresponding oxalate
18 anion [31]. Some other peaks appeared along the reaction indicating the presence of other
19 possible intermediates that the ConcIRT software could not identify, such as ethylene glycol
20 diformate and methoxyacetic acid (Figure 6). Complementarily, the chromatographic analyses
21 detected the production of a low concentration of methoxyacetic acid and a negligible amount of
22 acetic acid; whereas ethylene glycol was actually found along the reaction, which pointed out
23 that the pathway of degradation mainly progressed through the formation of ethylene glycol
24 diformate.

1 As a consequence, a mechanism for the degradation pathway that was followed along the
2 Fenton treatment of 1,4-dioxane may be proposed based on a radical reaction mechanism
3 (Schemes 1-3). First, hydroxyl radicals that were generated by the reaction between Fe^{+2} and
4 H_2O_2 , attacked 1,4-dioxane molecules to form 1,4-dioxanyl radical. Thereafter, this reaction
5 proceeded until 1,4-dioxan- α -oxyl radical was obtained by the reaction of 1,4-dioxanyl radical
6 with available dissolved oxygen (Scheme 1), first resulting in the formation of the peroxy
7 radical that next underwent termination reactions to generate the tetroxide precursor of 1,4-
8 dioxan- α -oxyl radical, which was finally produced releasing oxygen [2,3,13]. Thereafter, 1,4-
9 dioxan- α -oxyl radical progressed splitting the C-C bond, which was stabilized by ring opening.
10 This linear peroxy radical evolved then reacting with oxygen to produce a linear tetroxide that
11 may undergo two further reactions: (1) the electrocyclic process of this linear tetroxide, which
12 finally yields ethylene glycol diformate [2,13]; or (2) the decomposition in two alkoxy radicals
13 and oxygen, followed by β -fragmentation and H abstraction, finally producing ethylene glycol
14 monoformate [13] (Scheme 1).

15 Stephan and Bolton [13] also proposed an alternative degradation pathway from 1,4-
16 dioxan- α -oxyl radical consisting on the formation of methoxyacetic acid, and finally resulting in
17 the formation of acetic acid. Although some peaks that were recorded along the reaction could be
18 assigned to methoxyacetic acid (Figure 6), and chromatographic analyses showed this by-
19 product was somewhat produced, the almost completely absence of acetic acid along the reaction
20 actually denotes that this degradation alternative was not significantly followed in comparison to
21 the degradation pathway through ethylene glycol. Nevertheless, Beckett and Hua [2] reported
22 that the degradation of methoxyacetic acid may also result in the generation of glycolic acid and
23 formic acid.

1 Finally, ethylene glycol mono- and diformate may have progressed by acid hydrolysis to
2 yield ethylene glycol and formic acid as by-products [32] (Scheme 2). Whereas ethylene glycol
3 may be degraded to glycolic acid, and then evolve to oxalic acid by deformilation [33]; oxalic
4 acid itself really shows up as oxalate anion under the acidic pH values predominating in the
5 solution (Scheme 3).

6 In summary, during the oxidation of 1,4-dioxane, several by-products were produced as
7 the consequence of the ring opening of 1,4-dioxane molecules, namely: ethylene glycol, glycolic
8 acid, oxalate anion and formic acid. As a result, all 1,4-dioxane was removed, resulting in the
9 appearance of more biodegradable by-products, which may allow a further biological treatment
10 stage when necessary. An 80% total COD removal was finally achieved during these trials
11 thanks to the previously identified optimum reaction conditions for this treatment and the further
12 selection of a continuous addition mode for H₂O₂ during the process, which has previously been
13 reported to address a similar treatment efficiency enhancement [18, 19].

14

15 **4. Conclusions**

16 The treatment of 1,4 dioxane by Fenton's reagent achieved the total removal of this compound
17 from wastewater, as well as 80% COD reduction efficiency under the best tested treatment
18 conditions. In addition, wastewater biodegradability may be further enhanced, which has been
19 addressed by a significant high presence of volatile fatty acids at the end of the reaction.

20 Therefore, although the total mineralization of 1,4-dioxane was not achieved, final persisting by-
21 products were highly biodegradable; thus allowing further conventional treatment if final
22 disposal requirements make it necessary.

23 FTIR technique allowed the identification of the main compounds involved in the
24 degradation of 1,4-dioxane by conventional Fenton treatment, as well as the control of the

1 behavior of the target compound, denoting its presence and evolution along the reaction. This
2 methodology also allowed addressing a new degradation mechanism, monitoring the formation
3 of by-products, and optimizing the process identifying the presence of biodegradable compounds
4 that may easily be treated by conventional biological technologies, which may furthermore
5 represent a saving in the overall treatment cost.

6

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11

12 REFERENCES

- 13 [1] P. Ghosh, A.N. Samanta, S. Ray, Oxidation kinetics of degradation of 1,4-dioxane in aqueous
14 solution by H₂O₂/Fe(II) system, *J. Environ. Sci. Health, Part A: Toxic/Hazard. Subst. Environ.*
15 *Eng.* 45 (2010) 395-399.
- 16 [2] M.A. Beckett, I. Hua, Enhanced sonochemical decomposition of 1,4-dioxane by ferrous iron,
17 *Water Res.* 37 (2003) 2372-2376.
- 18 [3] H.S. Kim, B.H. Kwon, S.J. Yoa, I.K. Kim, Degradation of 1,4-Dioxane by Photo-Fenton
19 Processes, *J. Chem. Eng. of Jpn.* 41, 8 (2008) 829-835.
- 20 [4] M.H. So, J.S. Han, T.H. Han, J.W. Seo, C.G. Kim, Decomposition of 1,4-dioxane by photo-
21 Fenton oxidation coupled with activated sludge in a polyester manufacturing process, *Water Sci.*
22 *Technol.* 59, 5 (2009) 1003-1009.

- 1 [5] J.H. Suh, M. Mohseni, A study on the relationship between biodegradability enhancement
2 and oxidation of 1,4-dioxane using ozone and hydrogen peroxide, *Water Res.* 38 (2004) 2596-
3 2604.
- 4 [6] F. Harber, J.J. Weiss, The catalytic decomposition of Hydrogen Peroxide by iron salts, *J.*
5 *Amer. Chem. Soc.* 45 (1934) 338-351.
- 6 [7] C.P. Huang, C. Dong, Z. Tang, Advanced chemical oxidation: its present role and potential
7 future in hazardous waste treatment, *Waste Manag.* 13 (1993) 361-377.
- 8 [8] J.J. Pignatello, E. Oliveros, A. MacKay, Advanced oxidation processes for organic
9 contaminant destruction based on the Fenton reaction and related chemistry, *Crit. Rev. Environ.*
10 *Sci. Technol.* 36 (2006) 1-84.
- 11 [9] D. Hermosilla, M. Cortijo, C.P. Huang, The role of iron on the degradation and
12 mineralization of organic compounds using conventional Fenton and photo-Fenton processes,
13 *Chem. Eng. J.* 155 (2009) 637-646.
- 14 [10] R.F.F. Pontes, J.E.F. Moraes, A. Machulek Jr., J.M. Pinto, A mechanistic kinetic model for
15 phenol degradation by the Fenton process, *J. Hazard. Mater.* 176 (2010) 402-413.
- 16 [11] E. Khan, W. Wirojanagud, N. Sermsai, Effects of iron type in Fenton reaction on
17 mineralization and biodegradability enhancement of hazardous organic compounds, *J. Hazard.*
18 *Mater.* 161 (2009) 1024-1034.
- 19 [12] H.S. Son, J.K. Im, K.D. Zoh, A Fenton-like degradation mechanism for 1,4-dioxane using
20 zero-valent iron (Fe⁰) and UV light, *Water Res.* 43 (2009) 1457-1463.
- 21 [13] M.I. Stefan, J.R. Bolton, Mechanism of the Degradation of 1,4-Dioxane in Dilute Aqueous
22 Solution Using the UV/Hydrogen Peroxide Process, *Environ. Sci. Technol.* 32 (1998) 1588-
23 1595.

- 1 [14] V. Guzsavány, L. Rajic, B. Jovic, D. Orcic, J. Csanádi, S. Lazic, B. Abramovic,
2 Spectroscopic monitoring of photocatalytic degradation of the insecticide acetamiprid and its
3 degradation product 6-chloronicotinic acid on TiO₂, J. Environ. Sci. Health, Part A:
4 Toxic/Hazard. Subst. Environ. Eng. 47 (2012) 1919-1929.
- 5 [15] V. Guzsavány, M. Kádár, Z. Papp, L. Bjelica, F. Gaál, K. Tóth, Monitoring of Photocatalytic
6 Degradation of Selected Neonicotinoid Insecticides by Cathodic Voltammetry with a Bismuth
7 Film Electrode, Electroanalysis 20 (2008) 291 – 300.
- 8 [16] APHA, AWWA, WPCF (Eds.), Standard methods for the examination of water and
9 wastewater, Washington DC, 2005.
- 10 [17] H. Pobiner, Determination of hydroperoxides in hydrocarbon by conversion to hydrogen
11 peroxide and measurement by titanium complexing, Anal. Chem. 33 (1961) 1423-1428.
- 12 [18] D. Hermosilla, M. Cortijo, C.P. Huang, Optimizing the treatment of landfill leachate by
13 conventional Fenton and photo-Fenton processes, Sci. Total Environ. 407 (2009) 3473-3481.
- 14 [19] J.A. Zazo, J.A. Casas, A.F. Mohedano, M.A. Gilarranz, J.J. Rodríguez, Chemical Pathway
15 and Kinetics of Phenol Oxidation by Fenton's Reagent, Environ, Sci. Technol. 39 (2005) 9295-
16 9302.
- 17 [20] C.G. Kim, H.J. Seo, B.R. Lee, Decomposition of 1,4-Dioxane by Advanced Oxidation and
18 Biochemical Process, J. Environ. Sci. Health, Part A: Toxic/Hazard. Subst. Environ. Eng. 41
19 (2006) 599-611.
- 20 [21] R.L. Mason, R.F. Gunst, J.L. Hess, (Eds.), Statistical design and analysis of experiments,
21 eight applications to engineering and science, 2nd ed., Wiley: New York, 2003.
- 22 [22] Y. Wu, S. Zhou, F. Qin, X. Ye, K. Zheng, Modeling physical and oxidative removal
23 properties of Fenton process for treatment of landfill leachate using response surface
24 methodology (RSM), J. Hazard. Mater. 180 (2010) 456-465.

- 1 [23] D. Hermosilla, N. Merayo, R. Ordoñez, A. Blanco, Optimization of conventional Fenton
2 and ultraviolet-assisted oxidation processes for the treatment of reverse osmosis retentate from a
3 paper mill, *Waste Manage.* 32 (2012) 1236-1243.
- 4 [24] H. Gulsen, M. Turan, Treatment of sanitary landfill leachate using a combined anaerobic
5 fluidized bed reactor and Fenton's oxidation, *Environ. Eng. Sci.* 21 (2004) 627-636.
- 6 [25] H. Zhang, H.J. Choi, C.P. Huang, Optimization of Fenton process for the treatment of
7 landfill leachate, *J. Hazard. Mater.* 125 (2005) 166-174.
- 8 [26] J. Kiwi, C. Pulgarin, P. Peringer, M. Gratzel, Beneficial-effects of homogeneous photo-
9 Fenton pretreatment upon the biodegradation of anthraquinone sulfonate in waste-water
10 treatment, *Appl. Catal. B* 3 (1993) 85-99.
- 11 [27] C. Pulgarin, J. Kiwi, Overview on photocatalytic and electrocatalytic pretreatment of
12 industrial non-biodegradable pollutants and pesticides, *Chimia* 50 (1996) 50-55.
- 13 [28] Y.W. Kang, K. Hwang, Effects of reaction conditions on the oxidation efficiency in the
14 Fenton process, *Water Res.* 34 (2000) 2786-2790.
- 15 [29] D. Hermosilla, N. Merayo, R. Ordoñez, A. Blanco, Optimization of conventional Fenton
16 and ultraviolet-assisted oxidation processes for the treatment of reverse osmosis retentate from a
17 paper mill, *Waste Manage.* 32 (2012) 1236-1243.
- 18 [30] R.T.S.M. Lakshmi, M.K. Vyas, A.S. Brar, I.K. Varma, Synthesis and characterization of
19 sulphonated PEES copolymers by NMR spectroscopy, *Eur. Polym. J.* 42 (2006) 1423-1432.
- 20 [31] N. Merayo, D. Hermosilla, C. Negro, A. Blanco, On-line FTIR as a novel tool to monitor
21 Fenton processes behaviour, *Chem. Eng. J.* 232 (2013) 519-526.
- 22 [32] K.P.C. Vollhardt, N.E. Schore, *Organic Chemistry: Structure and Function*, 6th Ed., W.H.
23 Freeman and Co., Reston (VA), USA, 2011.

[33] V.Maurino, P.Calza, C.Minero, E.Pehzzetti, M.Vincenti, Light assisted 1,4-dioxane degradation, Chemosphere 35 (1997) 2675-2688.

Table 1. Estimated regression coefficients and standard errors for quadratic model used in RSM analysis.

	Coefficient	Standard error
Constant	39.509	3.851
pH (X1)	-5.489	1.671
[H ₂ O ₂] ₀ /COD ₀ (X2)	11.503	1.656
[H ₂ O ₂] ₀ /[Fe ²⁺] ₀ (X3)	-6.046	1.669
X1 ²	4.000	2.866
X2 ²	-2.625	3.283
X3 ²	-4.550	2.908
X1 · X2	-2.679	1.990
X2 · X3	0.083	1.986
X1 · X3	3.836	2.022

1 **Figure captions**

2

3 **Figure 1.** Reference spectra for the main chemicals that are expected to be found along the
4 Fenton oxidation treatment of 1,4-dioxane.

5

6 **Figure 2.** Response surface and contour plot for the reduction of the COD resulting in the
7 Fenton treatment of 1,4-dioxane ($247.8 \text{ mg} \cdot \text{L}^{-1}$) being performed at different pH and reagents
8 concentrations values ($T = 25^\circ\text{C}$).

9

10 **Figure 3.** Pareto chart including the degree of influence of each considered factor in the Fenton
11 treatment of 1,4-dioxane.

12

13 **Figure 4.** Evolution of FTIR spectra within the $2000\text{-}750 \text{ cm}^{-1}$ region along the Fenton treatment
14 of 1,4-dioxane. *Reaction conditions:* $[1,4\text{-dioxane}] = 70 \text{ mmol}$, $\text{pH} = 2.8 \pm 0.2$, $[\text{H}_2\text{O}_2]_0 / [\text{Fe}^{2+}]_0 = 5$.

15

16 **Figure 5.** FTIR-absorbance profiles of the main identified by-products that are produced during
17 the Fenton oxidation of 1,4-dioxane. *Reaction conditions:* $[1,4\text{-dioxane}] = 70 \text{ mmol}$, $\text{pH} = 2.8 \pm 0.2$,
18 $[\text{H}_2\text{O}_2]_0 / [\text{Fe}^{2+}]_0 = 5$.

19

20 **Figure 6.** FTIR-absorbance profiles representing the evolution of representative peaks along the
21 degradation of 1,4-dioxane by Fenton's reagent within the spectral region of $2000\text{-}750 \text{ cm}^{-1}$ (989
22 cm^{-1} : ethylene glycol and methoxyacetic acid; 1631 cm^{-1} : glycolic acid, oxalate anion, and oxalic
23 acid; 1581 cm^{-1} : oxalate anion; 1380 cm^{-1} : 1,4-dioxane, ethylene glycol diformate, and formic

1 acid; 1715 cm^{-1} : ethylene glycol diformate, methoxyacetic acid, and formic acid). *Reaction*
2 *conditions*: [1,4-dioxane]=70 mmol, pH=2.8±0.2, $[\text{H}_2\text{O}_2]_0/[\text{Fe}^{2+}]_0=5$.

3

4 **Scheme 1.** Degradation mechanism generating ethylene glycol mono- and diformate from 1,4-
5 dioxane

6

7 **Scheme 2.** Degradation route for the acidic hydrolysis of ethylene glycol diformate into ethylene
8 glycol and formic acid.

9

10 **Scheme 3.** Degradation route producing glycolic and oxalic acid from ethylene glycol.

FIGURE 1

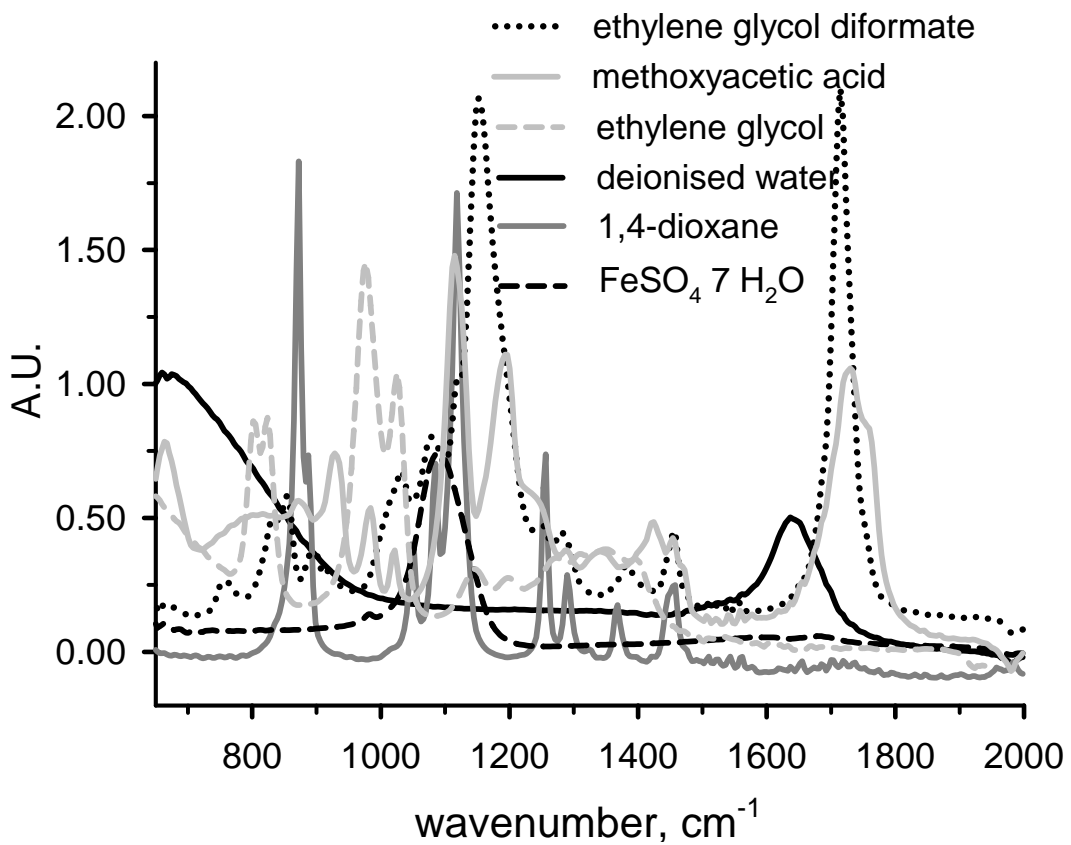
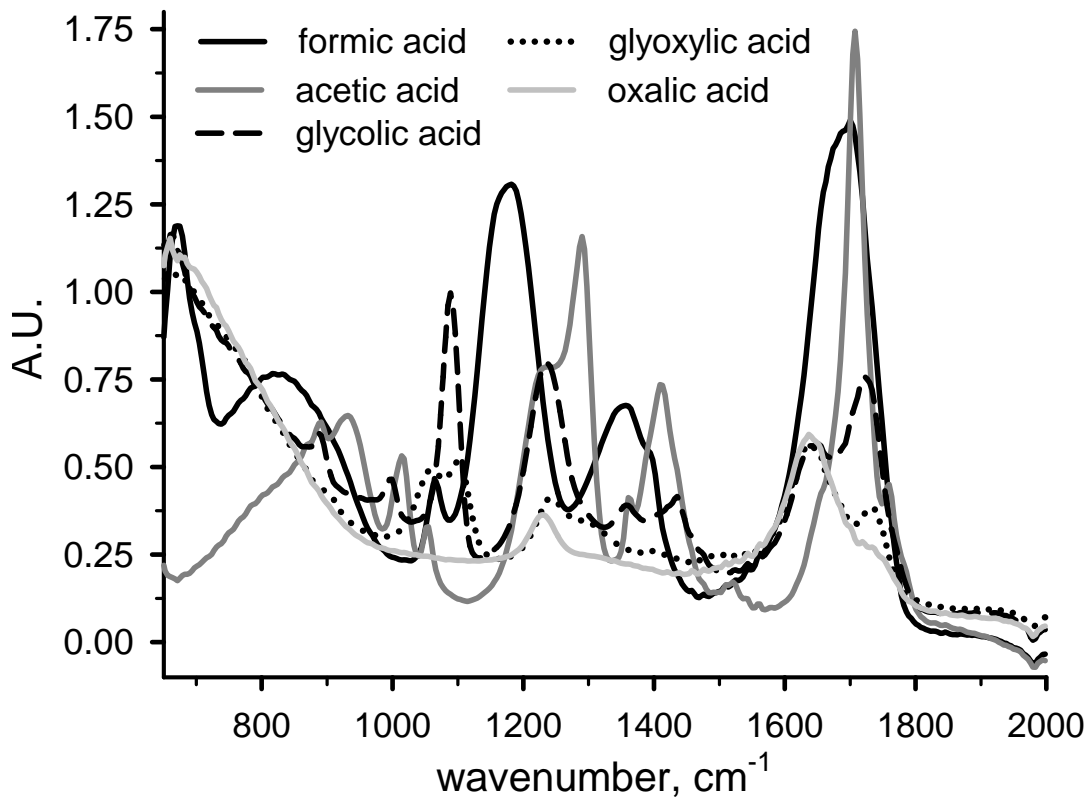


FIGURE 2

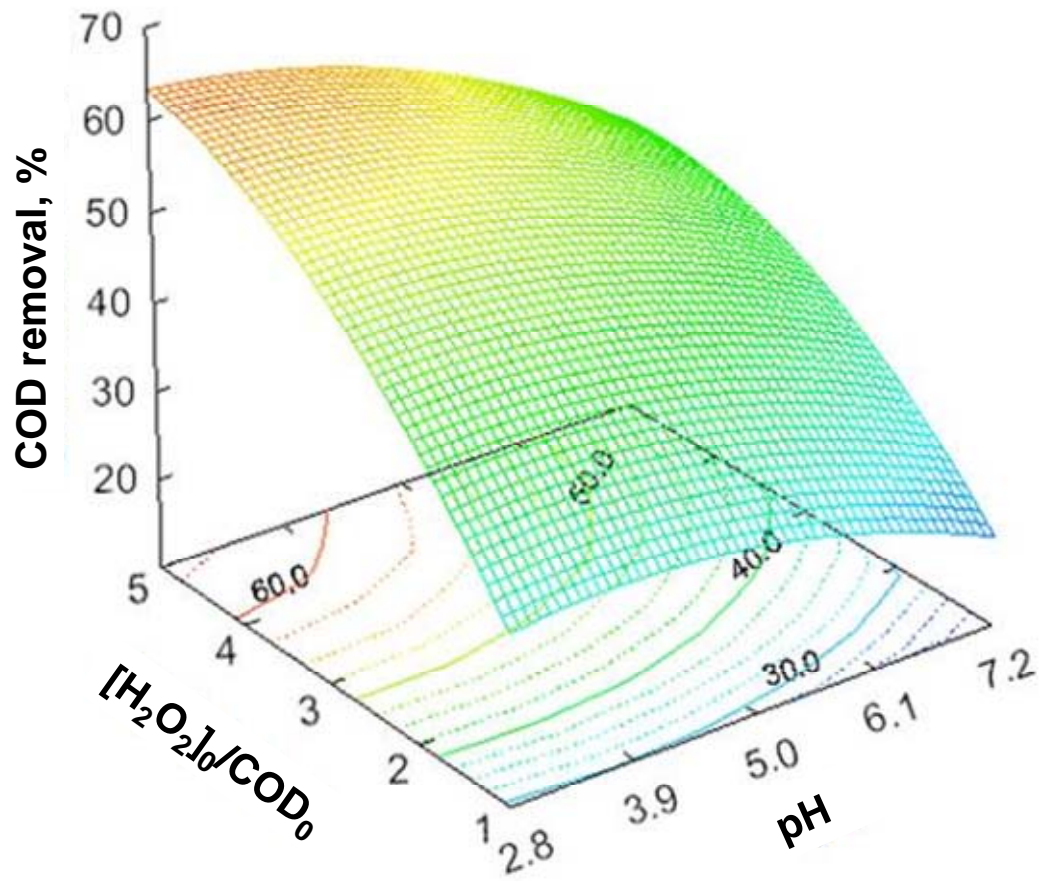


FIGURE 3

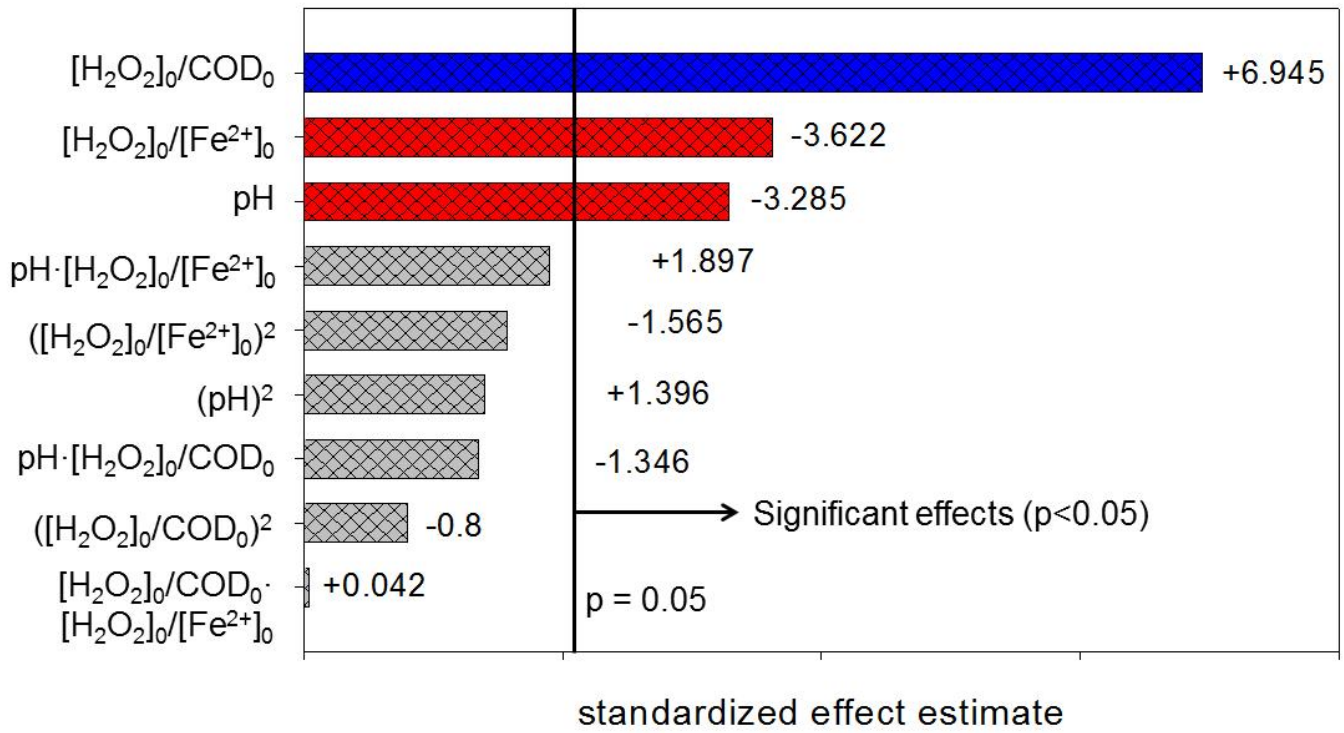


FIGURE 4

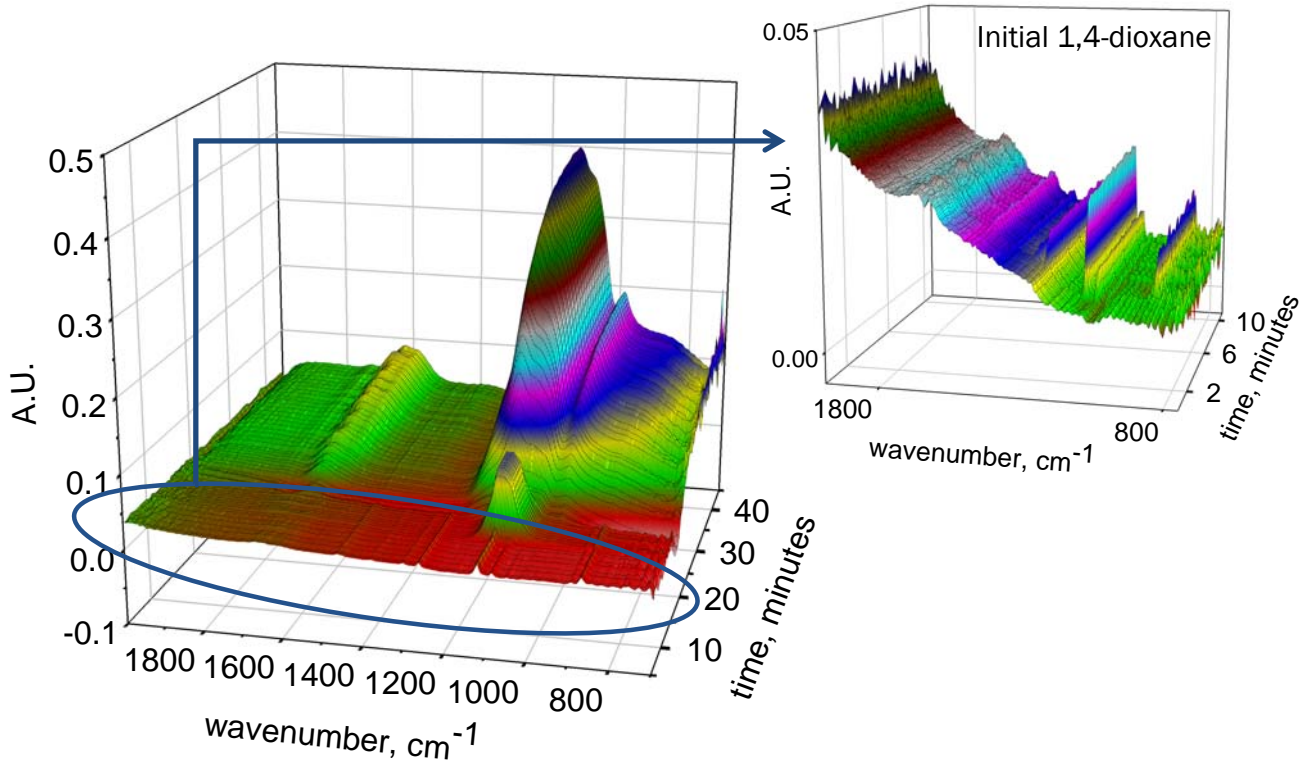


FIGURE 5

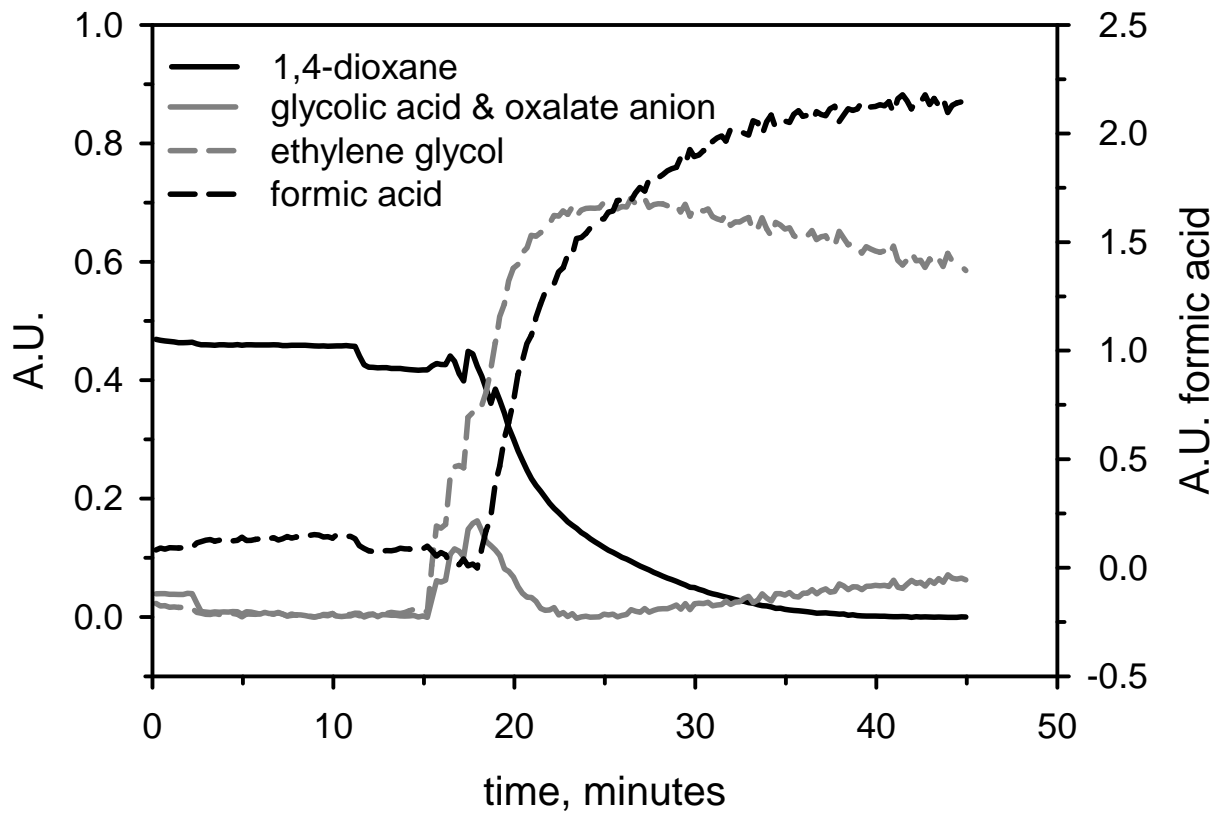
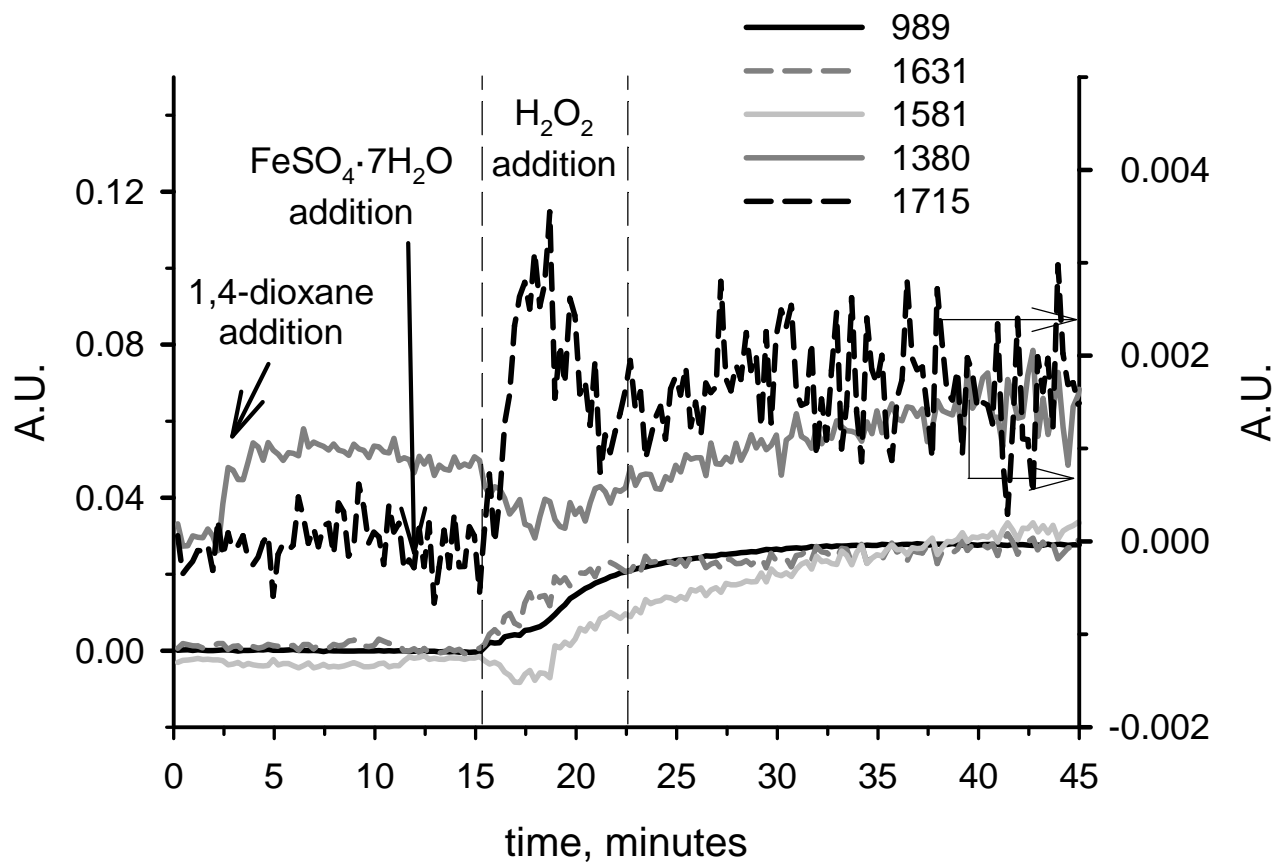
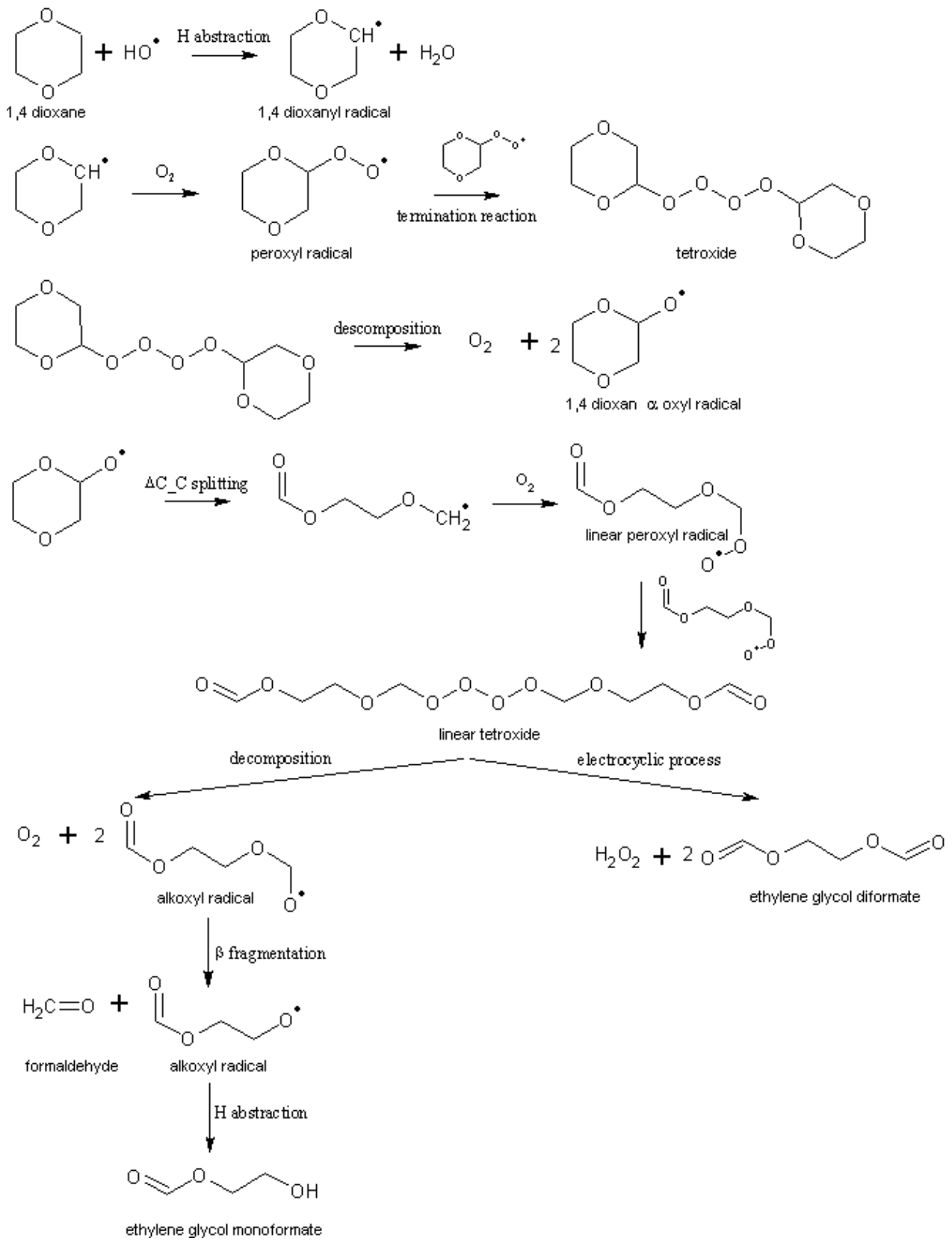


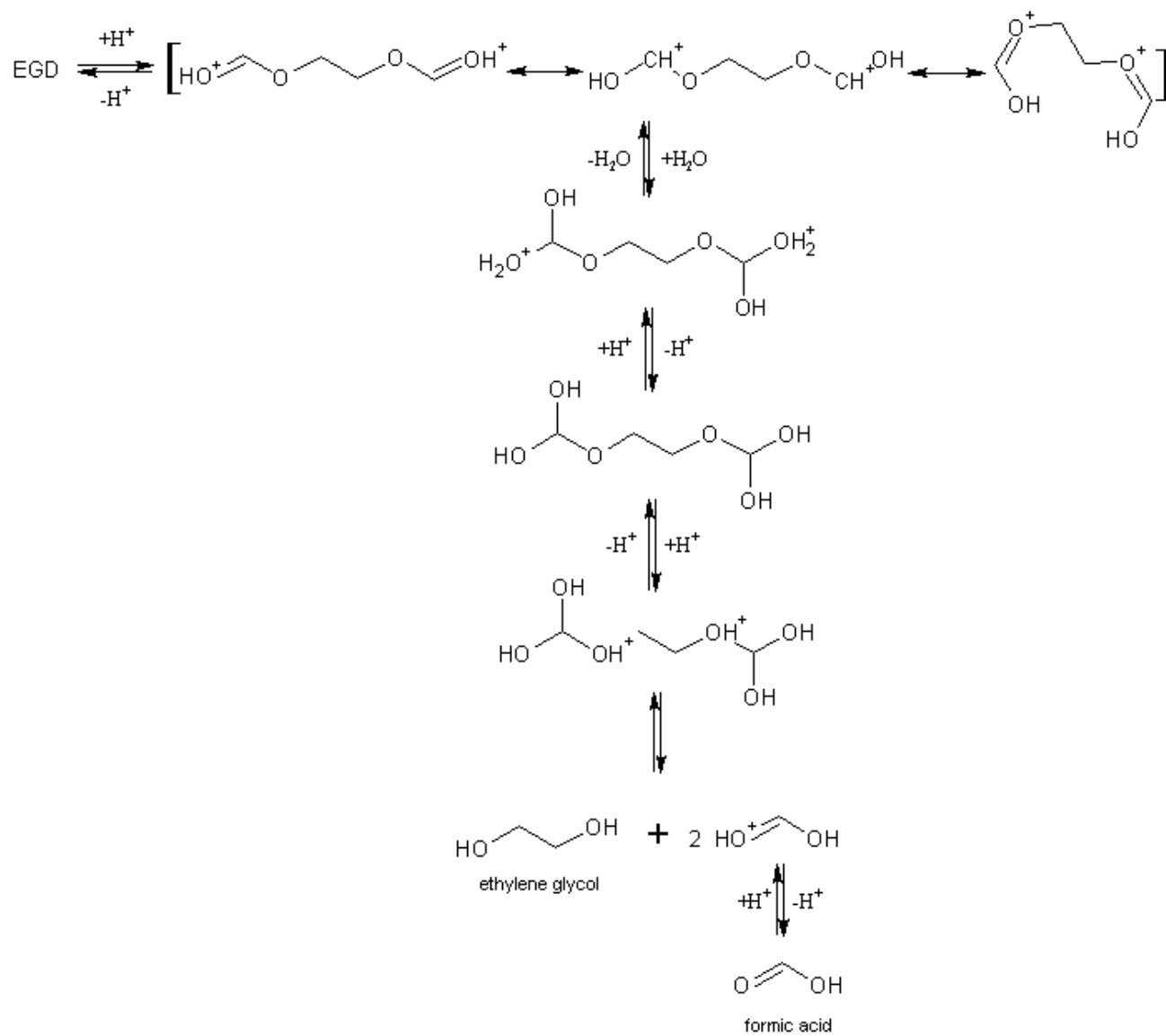
FIGURE 6



SCHEME 1



SCHEME 2



SCHEME 3

